

Research report

Assessment of luteolin (3',4',5,7-tetrahydroxyflavone) neuropharmacological activity

Miguel Coleta^{a,*}, Maria Graça Campos^a, Maria Dulce Cotrim^b,
Thereza Christina M. de Lima^c, António Proença da Cunha^a

^a Laboratory of Pharmacognosy, Center for Pharmaceutical Studies, University of Coimbra, 3000 Coimbra, Portugal

^b Laboratory of Pharmacology, Faculty of Pharmacy, University of Coimbra, 3000 Coimbra, Portugal

^c Laboratory of Neuropharmacology, Department of Pharmacology, CCB, Universidade Federal de Santa Catarina, Florianópolis, 88039-900, SC, Brazil

Received 11 October 2007; received in revised form 10 December 2007; accepted 13 December 2007

Available online 23 December 2007

Abstract

Since the discovery that certain flavonoids (namely flavones) specifically recognise the central BDZ receptors, several efforts have been made to identify naturally occurring GABA_A receptor benzodiazepine binding site ligands. Flavonoid derivatives with a flavone-like structure such as apigenin, chrysin and wogonin have been reported for their anxiolytic-like activity in different animal models of anxiety. Luteolin (3',4',5,7-tetrahydroxyflavone) is a widespread flavonoid aglycon that was reported as devoid of specific affinity for benzodiazepine receptor (BDZ-R) binding site, but its psychopharmacological activity is presently unknown. Considering (1) the close structural similarity with other active flavones, (2) the activity of some of its glycosylated derivatives and (3) the complexity of flavonoid effects in the central nervous system, luteolin was submitted to a battery of tests designed to evaluate its possible activity upon the CNS and its ability to interact with the BDZ-receptor binding sites was also analysed.

Luteolin apparently has CNS activity with anxiolytic-like effects despite the low affinity for the BDZ-R shown *in vitro*. Our findings suggest a possible interaction with other neurotransmitter systems but we cannot rule out the possibility that luteolin's metabolites might show a higher affinity for the BDZ-R *in vivo*, thus eliciting the evident anxiolytic-like effects through a GABAergic mechanism.

© 2007 Elsevier B.V. All rights reserved.

Keywords: Luteolin; Flavonoids; Flavones; Central nervous system; Anxiolytics; GABA

1. Introduction

Flavonoids are a large group of plant secondary metabolites that share a basic phenylbenzopyrone feature and are found in all vascular plants where they occur in several structurally and biosynthetically related classes [1]. They are important constituents of the human diet [2] and can also be found in expressive amounts in many medicinal plants [3]. Amongst the wide range of biological and pharmacological properties of these compounds we find a series of reports on their activity in the central nervous system (CNS) (for reviews see [4–6]). Since the discovery that certain flavonoids (namely flavones) specifically

recognise the central BDZ receptors [7,8], efforts have been made to identify naturally occurring GABA_A receptor benzodiazepine binding site ligands [5] to understand their interaction with these receptors [9–12] and to establish the CNS activity of different natural [13] and synthetic flavonoids [14,15]. Amongst these reports flavonoid derivatives with a flavone-like structure such as apigenin [16,17], chrysin [18] and wogonin [19] have been reported for their anxiolytic-like activity in different animal models of anxiety. These flavonoids with BDZ-receptor specificity and/or anxiolytic activity have been isolated from medicinal plants traditionally used in folk medicine for their anxiolytic/sedative properties such as *Passiflora coerulea* [20], *Matricaria recutita* [16], *Tilia tomentosa* [21], *Jatropha cillolata* [22], *Salvia guaranitica* [23], *Matricaria chamomilla* [17], *Ziziphus jujuba* [24]. Recently, we have reported on the isolation of luteolin-7-*O*-(2-rhamnosyl)glucoside from *Passiflora edulis* Sims and demonstrated its anxiolytic-like activity [25].

* Corresponding author at: R. dos Girassóis, 258, 2785-725 Cascais, Portugal. Tel.: +351 917600504; fax: +351 239827126.

E-mail address: mcoleta71@gmail.com (M. Coleta).

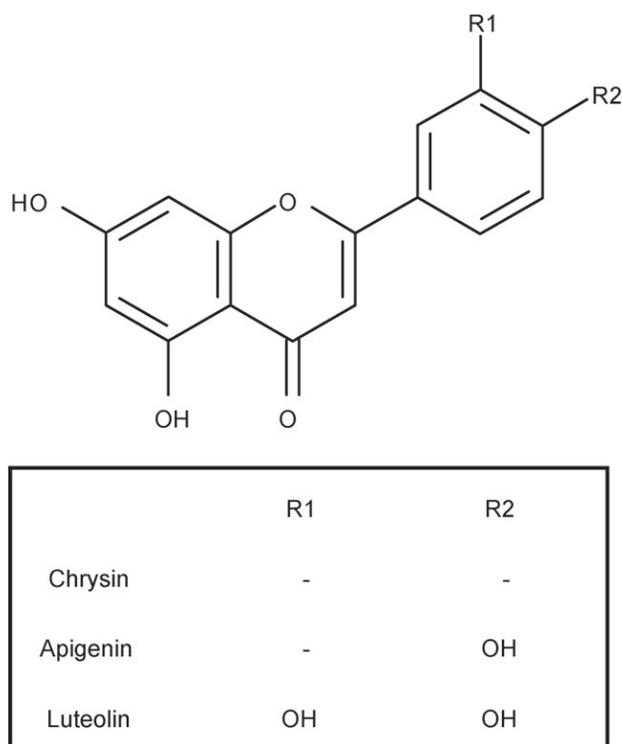


Fig. 1. Luteolin's structure is close to that of other flavones that have been reported for its anxiolytic activity like apigenin or chrysin.

Luteolin is a widespread flavonoid aglycon that was reported as devoid of specific affinity for BDZ-receptor binding site [21], but its psychopharmacological activity is presently unknown. Considering (1) the close structural similarity with other active flavones (Fig. 1), (2) the activity of some of its glycosylated derivatives [25,26,22] and (3) that flavonoid effects in the central nervous system are complex and can involve different mechanisms [27] besides the interaction with the benzodiazepine binding sites (BDZ-bs) at the GABA_A receptors, we became interested in the possible psychopharmacological profile of action of luteolin. This substance, purchased from a commercial source, was submitted to a battery of tests designed to evaluate its possible activity upon the CNS and to an eventual understanding of mechanisms underlying its activity(ies). As we were also interested in analysing the ability of luteolin to interact with the BDZ-receptor binding sites, we have also evaluated this substance in a radioreceptor binding assay with [³H]flunitrazepam.

2. Material and Methods

2.1. Animals

Male adult Swiss mice from our breeding stock, weighing 20–25 g, were used. Animals were placed in groups of 10 with free access to water and food, except during the experiments. They were kept on a 12/12 h day/night cycle (lights on at 07:00 a.m.) at controlled room temperature (23 ± 2 °C) and were allowed to adapt to the laboratory conditions for, at least, 1 week before the beginning of the behavioral experiments. Each animal was used just once. All experiments were conducted in accordance with international standards of animal welfare recommended by the Brazilian Society of Neuroscience and Behavior. The experimental protocols were approved by the local Animal Care

and Use Committee. All efforts were made to minimize animal suffering and to reduce the number of animals used and all behavioral testing was performed during the animal's day light period between 09:00 a.m. and 01:00 p.m.

2.2. Drugs

Diazepam i.v. solution (Dienpax ®, Sanofi-Winthrop Lab., Brazil) was diluted with distilled water and used in the dose of 1 mg/kg as reference drug (positive control) for anxiolytic, sedative, muscle relaxant and anticonvulsant activities. Luteolin, the flavonoid compound, was purchased from Extrasynthèse (Genay, France). [³H]flunitrazepam was obtained from Amersham Biosciences.

2.3. Treatments

Luteolin was freshly suspended (in an ultrasound bath) in a suitable amount of distilled water to be acutely (1 h) or repeatedly (14 days) administered *per os* (*p.o.*) by an intragastric cannula. Doses of luteolin (0.1–50 mg/kg) as well as the time intervals were determined in preliminary tests. Control groups received only distilled water in equivalent volumes by the same route. The behavioral tests were performed in a soundproof room between 09:00 a.m. and 01:00 p.m. to reduce the confounding influence of diurnal variation in spontaneous behavior.

2.4. Procedures

2.4.1. Motor performance evaluation

Muscle relaxant effects were evaluated using the horizontal-wire test that consists of a stretched copper wire placed 20 cm above the ground [28]. Motor coordination was assessed using a rota-rod apparatus. This equipment has a 2.5 cm bar, rotating at 12 rpm, divided in six parts and placed at a height of 25 cm. Latency to fall from the rotating bar and number of falls in a period of 1 min test were registered [29].

2.4.2. Elevated plus-maze test (EPM)

The elevated plus-maze was slightly modified from that used by Lister [30]. Briefly, it consisted of two open arms (30 cm × 5 cm × 0.25 cm) and two enclosed arms (30 cm × 5 cm × 15 cm), extending from a central platform (5 cm × 5 cm) and raised 50 cm above floor level. The maze floor was constructed from black Plexiglas and the walls from clear Plexiglas. The conventional spatial-temporal measures recorded were the number of entries (all four paws on open or enclosed arms and expressed as percentage of total entries), the time spent on open arms (expressed as percentage of time spent on closed plus open arms), number of entries on enclosed arms and the time on the central platform. Ethologically derived measures were grooming, rearing, stretched attend postures (SAP), head-dipping (HD) and defecation as an emotionally related parameter [31]. A selective increase in the parameters of exploration of the open arms of the maze reveals an anxiolytic effect [32].

2.4.3. Hole-board test

The hole-board consisted of a square box made of transparent Plexiglas (50 cm × 50 cm × 30 cm), 10 cm above table surface, with equally distributed nine holes, 2 cm in diameter. The area of the hole-board is divided with white ink into 24 smaller areas. During 5 min we registered the number of head-dips, grooming behavior, rears and also of displacements between the different areas (locomotor activity) [33].

2.4.4. Potentiation of barbiturate-induced loss of righting reflex

One hour after treatment with luteolin, animals were administered (*i.p.*) with sodium pentobarbital (50 mg/kg). Latency for the loss of the righting reflex and its total duration was registered for three consecutive hours [34].

2.4.5. Catalepsy test

Animals' forepaws were placed over a horizontal glass tube standing 5 cm above floor surface, each 10 min interval for up 1 h. Catalepsy was evaluated as the time until removal of the forefeet from the tube. Two different sets of

experiments were performed: (1) animals were treated with 5, 10 and 50 mg/kg of pure luteolin and submitted to the test conditions; (2) animals previously treated with luteolin were administered with haloperidol (1 mg/kg) immediately before testing [35].

2.4.6. Maximal electroshock test

One hour after treatment, mice were submitted to a transcorneal electrical stimulation (50 mA; 0.2 s; 60 Hz). The flexion time (flexion of the front limbs) and the extension time (full hind limbs extension) of the convulsions elicited by the electrical stimulus, as well as the incidence and lethality of the induced convulsions, were registered [36].

2.4.7. Forced-swimming test

The test was slightly modified from that proposed by Porsolt et al. [37] and consisted in one exposure (6 min) to a water tank (height, 35 cm; diameter, 24 cm, with a water column of 13.5 cm at 25 °C). We have registered total immobilization time and the latency for this behavior after the first minute.

2.4.8. Rectal temperature evaluation

Body temperature was measured through a glycerin-lubricated thermistor (Lumiscop 2018) probe inserted about 1 cm into the rectum of the animal immediately before (basal values) and 1 h after treatment.

2.4.9. Statistical analysis

Data were analysed with Graphpad Prism® (v4.03). The statistical tests used were one-way ANOVA followed by Dunnett's test for comparison of treatment groups with control and Tukey's test for comparison between all treatment groups. Radioligand binding data were analysed by non-linear regression tools provided by the same software.

2.4.10. In vitro radioreceptor binding assay

Crude synaptic membranes were prepared from isolated rat brain cortices as previously described elsewhere [38].

Binding assays were performed using a semi-automatic filtration technique with diazepam (100 µM) to obtain the specific binding. Competition curves were obtained by adding to the assay tubes buffer solution (40 mM Hepes-Tris, pH 7.4), luteolin (5.55 µM–3.5 mM) or diazepam, followed by [³H]flunitrazepam (88 Ci/mmol—final concentration 1.5 nM in the inhibition curves and 0.014–20 nM in the saturation curves) and finally brain tissue homogenate (about 300–400 mg protein) was added to initiate binding. The assays were done in triplicate and tubes were incubated at 37 °C during 30 min and terminated by rapid filtration through glass fiber filters. The radioactivity remaining in the filters was determined in 8 ml of scintillation liquid (toluene 1 L, 167 mg of 2,5-difeniloxazol 7.3 g, *p*-bis(2(5-feniloxazoil(-benzene and 250 ml of Triton X-100) in a Packard Tri-Carb 2500 TR scintillation counter.

3. Results

3.1. Motor performance evaluation

Acute treatment with luteolin (1–50 mg/kg) did not affect the motor coordination or muscle relaxation of the animals, as measured on the rota-rod (ANOVA: $F_{4,38} = 1.705$; $p > 0.05$) and horizontal-wire tests (ANOVA: $F_{4,38} = 0.7528$; $p > 0.05$).

Also, there were no changes in the parameters directly related with motor activity in the hole-board test (number of crossings between the different sections) (ANOVA: $F_{4,38} = 0.8242$; $p > 0.05$) and in the elevated plus-maze (total number of entries in the closed arms of the maze: data not shown; ANOVA: $F_{4,38} = 1.311$; $p > 0.05$).

3.2. Anxiolytic activity

ANOVA showed a significant difference within treated groups ($F_{5,46} = 6.705$; $p < 0.0001$) and Dunnett's test revealed that only with the dose of 5 mg/kg there was a significant increase ($p < 0.05$) in the percentage of entries in the open areas of the EPM after acute treatment with luteolin (Fig. 2). As for the percentage of time spent in those areas we could not find significant differences between luteolin-treated groups and control groups ($F_{4,38} = 1.624$; $p > 0.05$). ANOVA analysis of ethological parameters namely unprotected head-dipping and stretch approach postures also revealed significant differences when compared with the control group ($F_{4,38} = 4.264$ and 3.486 , with $p = 0.0060$ and 0.0021 , respectively) and Dunnett's test showed there was a significant increase in unprotected head-dipping ($p < 0.05$) and a decrease in the stretch approach postures ($p < 0.05$) displayed after the administration of this same dose (Table 1).

3.3. Sedative activity

ANOVA showed a significant difference within treated groups ($F_{6,50} = 13.83$; $p < 0.0001$) and with Tukey's multiple comparison test we could observe a significant reduction of the latency time for the pentobarbital-induced loss of righting reflex, even with relatively low doses of luteolin (0.1 mg/kg; $p < 0.05$) and a more pronounced effect when doses higher

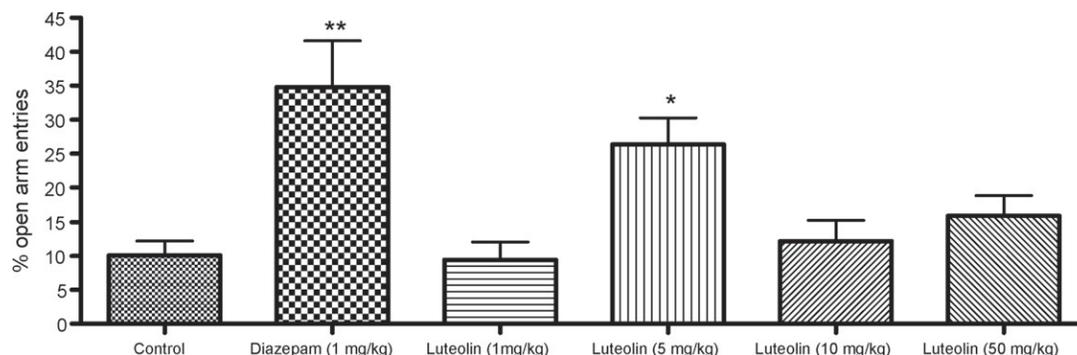


Fig. 2. Performance of mice in the elevated plus-maze after acute treatment with 1, 5, 10 and 50 mg/kg of luteolin. The percentage of entries in the open area was calculated with relation to the total number of entries in both closed and open arms of the maze. Results are expressed as mean \pm S.E.M. ($n = 8-10$). * $p < 0.05$; ** $p < 0.01$ versus control (vehicle treated group; Dunnett's test).

Table 1
Ethologically derived measures in the elevated plus-maze

	Control	Luteolin 1 mg/kg	Luteolin 5 mg/kg	Luteolin 10 mg/kg	Luteolin 50 mg/kg
Protected head-dipping	12.6 ± 3.7	13.98 ± 4.9	9.3 ± 2.9	9.7 ± 3.9	11.3 ± 4.8
Unprotected head-dipping	0.8 ± 0.7	0.4 ± 0.5	2.9 ± 0.6*	1.6 ± 1.5	0.8 ± 0.6
Rearing	22.8 ± 7.0	19.4 ± 5.5	27.7 ± 5.8	18.0 ± 4.0	20.8 ± 4.2
Immobility	0.6 ± 1.0	1.4 ± 1.8	1.7 ± 2.2	2.5 ± 1.5	6.0 ± 6.4
Grooming	4.2 ± 4.2	4.0 ± 3.7	0.7 ± 0.9	1.3 ± 1.3	4.6 ± 4.2
SAP	9.4 ± 2.5	5.5 ± 4.7	4.3 ± 1.1*	11.2 ± 3.4	12.3 ± 4.1

Results are expressed as mean ± S.E.M. ($n = 8-10$). * $p < 0.05$ versus control (vehicle treated group) (Dunnett's test).

than 1 mg/kg were tested (Fig. 3), as concluded from the fact that the effect observed after the administration of luteolin 0.1 mg/kg was significantly lower than after luteolin 5 and 10 mg/kg ($p < 0.05$). ANOVA analysis of the total duration of the pentobarbital-induced loss of righting reflex data showed significant differences between treatment groups ($F_{6,50} = 17.31$; $p < 0.0001$); however, the total duration was only significantly increased with high doses of luteolin (10 mg/kg) ($p < 0.05$, with Dunnett's test) (Fig. 4). Also, we have observed that during the loss of righting reflex all the animals treated with luteolin and, particularly, groups that received the dose of 5 and 10 mg/kg, showed unusual tremors of both anterior and posterior limbs.

3.4. Anticonvulsant activity

Compared with control groups, animals treated with luteolin showed no differences in the flexion and extension times as well as in the lethality of the electroshock-induced seizures for any of the doses tested (data not shown).

3.5. Catalepsy test

Luteolin (5, 10 and 50 mg/kg) did not induce catalepsy *per se*, but analysing the results with ANOVA and Dunnett's test we could conclude that the dose of 5 mg/kg significantly antagonised catalepsy induced by haloperidol ($F_{3,24} = 8.201$; $p < 0.01$)

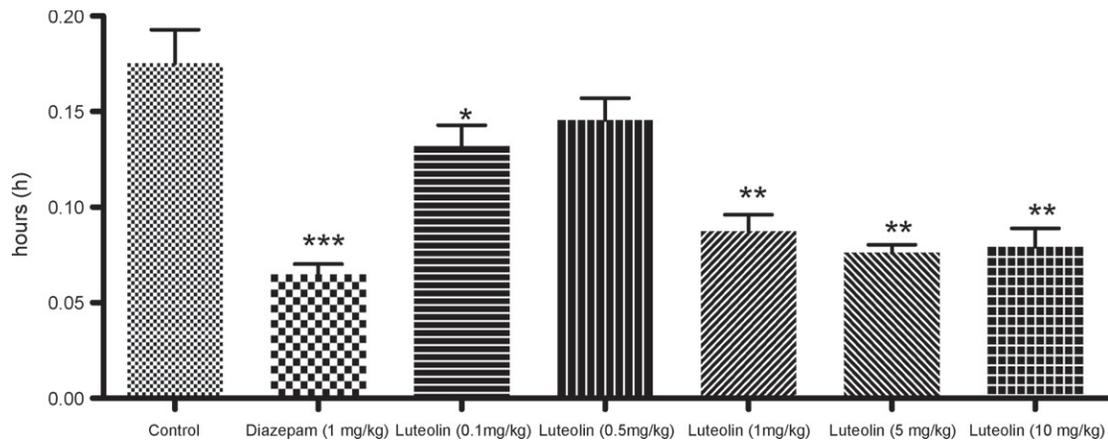


Fig. 3. Latency to the loss of the righting reflex induced by pentobarbital. Results are expressed as mean ± S.E.M. ($n = 7-11$). * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ versus control (vehicle treated group; Tukey's test).

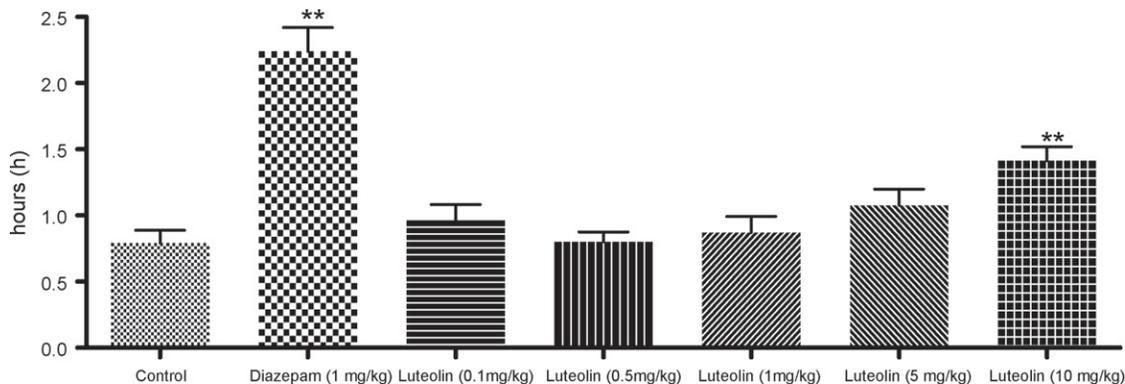


Fig. 4. Total duration of the loss of the righting reflex induced by pentobarbital. Results are expressed as mean ± S.E.M. ($n = 7-11$). ** $p < 0.01$ versus control (vehicle treated group; Dunnett's test).

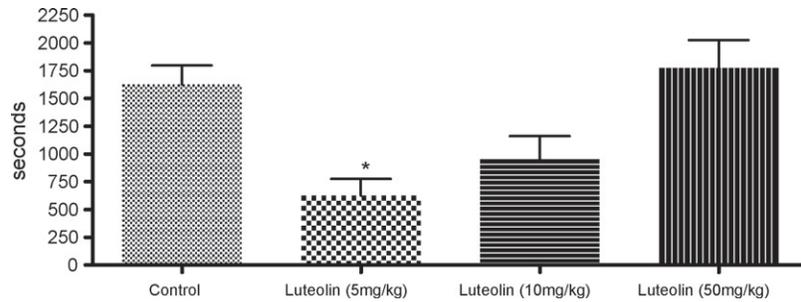


Fig. 5. Antagonism of haloperidol-induced catalepsy. Catalepsy was evaluated as the time of involuntary permanence of the animal in an unusual position (time until removal of the forefeet of the tube). Results are expressed as mean \pm S.E.M. ($n=6-8$). * $p < 0.01$ versus control (vehicle treated group; Dunnett's test).

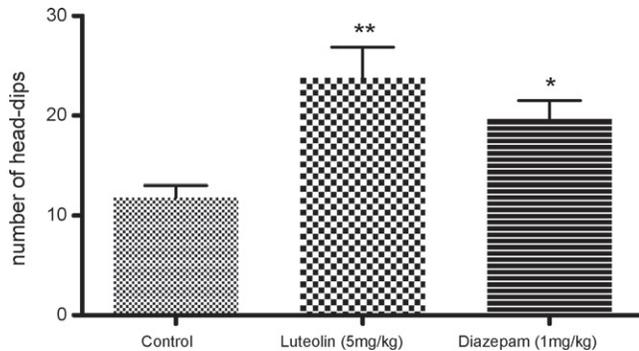


Fig. 6. Effect of chronic treatment with luteolin (5 mg/kg) on the frequency of head-dips in the hole-board test. Results are expressed as mean \pm S.E.M. ($n=9-11$). * $p < 0.05$; ** $p < 0.01$ versus control (vehicle treated group; Dunnett's test).

and this effect disappeared when higher doses (10, 50 mg/kg) ($F_{3,24} = 8.201$; $p > 0.05$) were tested (Fig. 5).

3.6. Chronic treatment

Animals treated with luteolin (5 mg/kg) daily for 14 days did not exhibit any significant changes either on the motor activity parameters as evaluated in the rota-rod ($F_{2,27} = 0.967$; $p > 0.05$) and horizontal-wire ($F_{2,27} = 0.822$; $p > 0.05$) tests or on the parameters related with motor activity in the EPM and hole-board test (data not shown).

Also, both ethological and spatio-temporal parameters of the EPM related with anxiolytic-like effects remained unchanged

after chronic treatment with luteolin (5 mg/kg); however, in the hole-board test, analysis of the results with ANOVA and Dunnett's test showed a significant increase in head-dipping ($F_{2,27} = 7.987$; $p < 0.01$; Fig. 6).

On the contrary, in the forced-swimming test, the ANOVA showed significant differences within treatment groups for both parameters measured ($F_{2,27} = 10.26$; $p = 0.0005$ and $F_{2,27} = 4.236$; $p = 0.0292$, for total immobilization time and latency to immobilization, respectively) and Dunnett's test revealed that repeated treatment with luteolin (5 mg/kg) significantly reduced the latency to immobilization ($p < 0.05$) and increased the total time of immobilization ($p < 0.01$; Fig. 7).

3.7. Radioreceptor binding assay

From [3 H]flunitrazepam saturation binding experiments K_d and B_{max} determined were 13.9 ± 3.3 nM and 10316 ± 1186 cpm, respectively. In competitive binding experiments carried out in the presence of 5 nM of [3 H]flunitrazepam, luteolin inhibited this radioligand binding to the rat cerebral cortex membranes (Fig. 8) with a K_i of $60.1 \mu\text{M}$ and a Hill slope of -0.91 .

4. Discussion

The results in the different tests show that luteolin after both acute and chronic treatment is devoid of muscle relaxant or motor

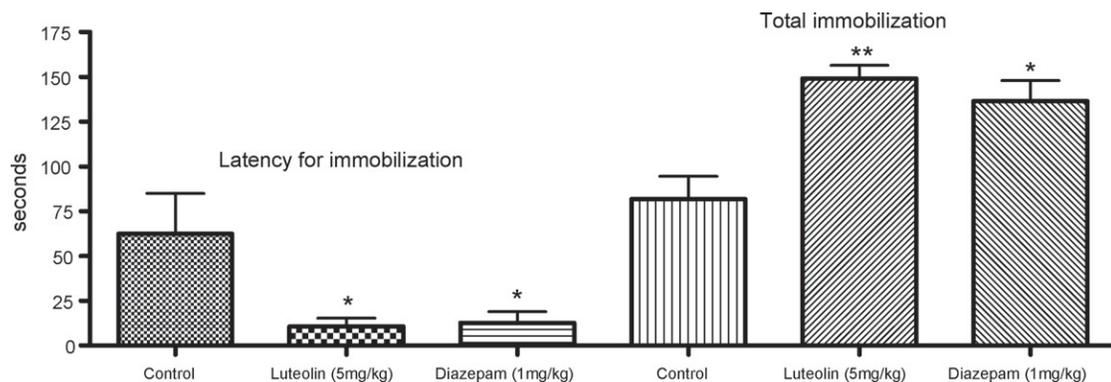


Fig. 7. Effect of chronic treatment with luteolin in the performance of mice in the forced-swimming test. We have registered total immobilization time and the latency to this behavior after the first minute. A mouse was considered to be immobile when it floated or made only small movements necessary to keep its head above water. Results are expressed as mean \pm S.E.M. ($n=9-11$). * $p < 0.05$; ** $p < 0.01$ versus control (vehicle treated group; Dunnett's test).

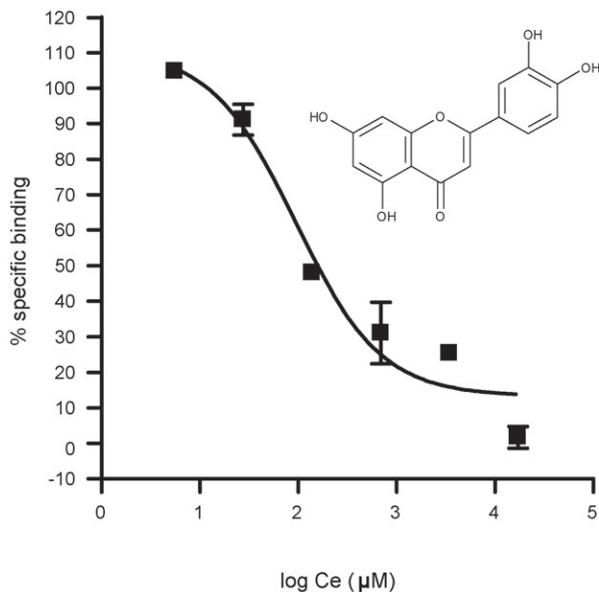


Fig. 8. Structure of luteolin and competitive inhibition curve of [³H]flunitrazepam binding to synaptosomal membranes.

coordination effects and activity on the CNS is thus not hindering motor activity performance.

Increased exploration in the open areas of the EPM (% open arm entries), diminution of SAP and increase in unprotected head-dipping are all consistent and suggest an anxiolytic-like effect [39]. However, one should expect the increased number of entries in the open areas of the maze also to reflect significantly in the percentage of time spent there; but that was not the case and, also, we could not observe any clear dose–effect relation in this test.

Potential of pentobarbital-induced loss of righting reflex can be elicited through interaction with different neurotransmitter systems, namely GABA [40,41] or 5-HT [42]. In our tests, the reduced latency time for the pentobarbital-induced loss of righting reflex elicited by luteolin (0.1, 5 and 10 mg/kg) and the increase in the total duration of this effect (10 mg/kg) can be interpreted as an indication of luteolin's possible interference with these systems.

Unlike classical benzodiazepines [43] and other flavone type BDZ-R ligands like apigenin [17] or chrysin [18], luteolin failed to give any protection against maximal electroshock.

Haloperidol is a potent D₂ antagonist [44] that elicits catalepsy. Besides dopamine, several neurotransmitters like serotonin, acetylcholine, GABA or endorphins are found to be involved in the expression of catalepsy and haloperidol-induced catalepsy can be blocked by such diverse drugs as selective dopamine D₃ receptor antagonists [45], 5-HT_{1A} agonists [46,47], anti-cholinergics [48] or A_{2A} receptor antagonists [49]. There are also well-known interactions between the GABAergic and dopaminergic system [50,51] and classic benzodiazepines like diazepam potentiate haloperidol-induced catalepsy [52]. On the contrary, GABA_A agonists like muscimol are reported to antagonise haloperidol-induced catalepsy at low doses with reverse effects at higher doses [53] and, interestingly enough,

GABA_B agonist baclofen antagonises the action of haloperidol in a low dose (1 mg/kg) without any visible effect in higher doses (2–8 mg/kg) [54]. There are reports of other flavonoid-type molecules like quercetin [55] or flavonoid-enriched extracts [56] interfering with haloperidol-induced catalepsy with different outcomes but the present results suggest that luteolin has a baclofen-like effect in this test. However, it must be noted that unlike baclofen [57], luteolin did not produce any significant reduction in the animals' body temperature (results not shown).

After the chronic treatment, our results of the hole-board test suggest an anxiolytic-like effect [58] but these were not observed in the EPM. Normally, the results with anxiolytics in both tests seem to correlate well but their sensitivity can differ [59] and that could explain these somewhat surprising results. On the contrary, it was previously reported that handling history can modify the behavioral effects of drugs in the EPM and GABA_B agonists like baclofen apparently exert an anxiolytic-like effect in this test only in handling naïve rats [60] and this could also explain the different results in the EPM after acute or repeated treatment. Moreover, the development of tolerance to the repeated treatment with luteolin could also explain its lack of effect in the EPM since tolerance is observed with the benzodiazepine drugs [61].

The meaning of immobility in swimming tests may vary in accordance with the protocol reflecting helplessness or adaptation in the forced-swimming test or in the swimming stress, respectively [62]. For mice, the forced-swimming conditions used in this test resemble more closely the situation of swimming-stress (once the animal does not touches the bottom with its hind paws) and in these conditions drugs like diazepam have the ability to increase immobility time [62,63] just as it was observed in our assay. Other structurally related compounds like apigenin, upon acute treatment with relatively high doses (25 mg/kg), have induced an antidepressant-like activity (reduction in immobility time) in the forced-swimming test [64].

4.1. Radioreceptor binding assay

Luteolin had previously been reported not to displace [³H]flunitrazepam binding to central benzodiazepine receptors (BDZ-R) (IC₅₀ > 100 µM) [21]. In our experiments we have determined that luteolin has in fact the ability to displace [³H]flunitrazepam binding, though exhibiting a low affinity for these receptors, with a K_i in the high (60.1) µM range. Despite the need to further analyse luteolin's interaction with BDZ-R, our results suggest that by itself this interaction does not seem to fully explain the results observed *in vivo*, thus prompting renewed interest in the analysis of possible interactions with other receptors.

Most of the literature published concerning the anxiolytic-like activity of flavone-type compounds has focused on the ability of these molecules to interact with the GABA_A benzodiazepine binding site (BDZ-bs) (for review see [5]). However, Luteolin, despite its low affinity for the BDZ-R, seems to have anxiolytic-like effects or, at least, to interact with different neurotransmitter systems so as to induce CNS effects. Another neurotransmitter system such as the 5-HT receptors [65,66]

could be involved in its action, and this should be further investigated. On the contrary, we must consider that flavonoids are subject to intense metabolism [67] and after oral administration of luteolin to rats, free luteolin has been determined in plasma but also luteolin's sulfate and glucuronate derivatives (the main metabolite was found to be a luteolin monoglucuronate) as well as *o*-methyl luteolin, with the dose administered strongly affecting the type of metabolites formed [68]. As there is no information about the affinity for the BDZ-receptor of luteolin's metabolites we cannot at this point discard the hypothesis that these might exhibit higher affinities for the BDZ-receptor, thus eliciting the evidenced anxiolytic-like effects through a GABAergic mechanism and this aspect should also be further investigated.

Acknowledgements

M. Coleta was a recipient of a PhD scholarship from Praxis XXI (BD/16264/98).

This work was performed during the B-national Cooperation Program in Research supported by GRICES (Portugal) and CNPq (Brazil).

References

- [1] Harborne JB, Williams CA. Advances in flavonoid research since 1992. *Phytochemistry* 2000;55(6):481–504.
- [2] Karakaya S, Sedef NEL. Quercetin, luteolin, apigenin and kaempferol contents of some foods. *Food Chem* 1999;66(3):289–92.
- [3] Middleton Jr E, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacol Rev* 2000;52(4):673–751.
- [4] Wang F, Shing M, Huen Y, Tsang SY, Xue H. Neuroactive flavonoids interacting with GABA_A receptor complex. *Curr Drug Targets CNS Neurol Disord* 2005;4(5):575–85.
- [5] Marder M, Paladini AC. GABA(A)-receptor ligands of flavonoid structure. *Curr Top Med Chem* 2002;2(8):853–67.
- [6] Paladini AC, Marder M, Viola H, Wolfman C, Wasowski C, Medina JH. Flavonoids and the central nervous system: from forgotten factors to potent anxiolytic compounds. *J Pharm Pharmacol* 1999;51(5):519–26.
- [7] Nielsen M, Frokjaer S, Braestrup C. High affinity of the naturally-occurring biflavonoid, amentoflavon, to brain benzodiazepine receptors in vitro. *Biochem Pharmacol* 1988;37(17):3285–7.
- [8] Medina JH, Pena C, Levi de Stein M, Wolfman C, Paladini AC. Benzodiazepine-like molecules, as well as other ligands for the brain benzodiazepine receptors, are relatively common constituents of plants. *Biochem Biophys Res Commun* 1989;165(2):547–53.
- [9] Huang X, Liu T, Gu J, Luo X, Ji R, Cao Y, et al. 3D-QSAR model of flavonoids binding at benzodiazepine site in GABA_A receptors. *J Med Chem* 2001;44(12):1883–91.
- [10] Dekermendjian K, Kahnberg P, Witt MR, Sterner O, Nielsen M, Liljefors T. Structure–activity relationships and molecular modeling analysis of flavonoids binding to the benzodiazepine site of the rat brain GABA(A) receptor complex. *J Med Chem* 1999;42(21):4343–50.
- [11] Marder M, Estiu G, Blanch LB, Viola H, Wasowski C, Medina JH, et al. Molecular modeling and QSAR analysis of the interaction of flavone derivatives with the benzodiazepine binding site of the GABA(A) receptor complex. *Bioorg Med Chem* 2001;9(2):323–35.
- [12] Hong X, Hopfinger AJ. 3D-pharmacophores of flavonoid binding at the benzodiazepine GABA(A) receptor site using 4D-QSAR analysis. *J Chem Inf Comput Sci* 2003;43(1):324–36.
- [13] Medina JH, Viola H, Wolfman C, Marder M, Wasowski C, Calvo D, et al. Overview—flavonoids: a new family of benzodiazepine receptor ligands. *Neurochem Res* 1997;22(4):419–25.
- [14] Wang F, Xu Z, Yuen CT, Chow CY, Lui YL, Tsang SY, et al. 6,2'-Dihydroxyflavone, a subtype-selective partial inverse agonist of GABA(A) receptor benzodiazepine site. *Neuropharmacology* 2007;53(4):574–82.
- [15] Griebel G, Perrault G, Tan S, Schoemaker H, Sanger DJ. Pharmacological studies on synthetic flavonoids: comparison with diazepam. *Neuropharmacology* 1999;38(7):965–77.
- [16] Viola H, Wasowski C, Levi de Stein M, Wolfman C, Silveira R, Dajas F, et al. Apigenin, a component of *Matricaria recutita* flowers, is a central benzodiazepine receptors-ligand with anxiolytic effects. *Planta Med* 1995;61(3):213–6.
- [17] Avallone R, Zanolli P, Puia G, Kleinschmitz M, Schreier P, Baraldi M. Pharmacological profile of apigenin, a flavonoid isolated from *Matricaria chamomilla*. *Biochem Pharmacol* 2000;59(11):1387–94.
- [18] Wolfman C, Viola H, Paladini A, Dajas F, Medina JH. Possible anxiolytic effects of chrysin, a central benzodiazepine receptor ligand isolated from *Passiflora coerulea*. *Pharmacol Biochem Behav* 1994;47(1):1–4.
- [19] Hui KM, Huen MS, Wang HY, Zheng H, Sigel E, Baur R, et al. Anxiolytic effect of wogonin, a benzodiazepine receptor ligand isolated from *Scutellaria baicalensis* Georgi. *Biochem Pharmacol* 2002;64(9):1415–24.
- [20] Medina JH, Paladini AC, Wolfman C, Levi de Stein M, Calvo D, Diaz LE, et al. Chrysin (5,7-di-OH-flavone), a naturally-occurring ligand for benzodiazepine receptors, with anticonvulsant properties. *Biochem Pharmacol* 1990;40(10):2227–31.
- [21] Viola H, Wolfman C, Levi de Stein M, Wasowski C, Peña C, Medina JH, et al. Isolation of pharmacologically active benzodiazepine receptor ligands from *Tilia tomentosa* (Tiliaceae). *J Ethnopharmacol* 1994;44(1):47–53.
- [22] Okuyama E, Okamoto Y, Yamazaki M, Satake M. Pharmacologically active components of a Peruvian medicinal plant, huanarpo (*Jatropha cillata*). *Chem Pharm Bull (Tokyo)* 1996;44(2):333–6.
- [23] Marder M, Viola H, Wasowski C, Wolfman C, Waterman PG, Medina JH, et al. Cirsiliol and caffeic acid ethyl ester, isolated from *Salvia guaranitica* are competitive ligands from the central benzodiazepine receptors. *Phytomedicine* 1996;3(1):29–31.
- [24] Peng WH, Hsieh MT, Lee YS, Lin YC, Liao J. Anxiolytic effect of seed of *Ziziphus jujuba* in mouse models of anxiety. *J Ethnopharmacol* 2000;72(3):435–41.
- [25] Coleta M, Batista MT, Campos MG, Carvalho R, Cotrim MD, Lima TC, et al. Neuropharmacological evaluation of the putative anxiolytic effects of *Passiflora edulis* Sims, its sub-fractions and flavonoid constituents. *Phytother Res* 2006;20(12):1067–73.
- [26] Fernández SP, Wasowski C, Loscalzo LM, Granger RE, Johnston GAR, Paladini AC, et al. Central nervous system depressant action of flavonoid glycosides. *Eur J Pharmacol* 2006;539(3):168–76.
- [27] Goutman JD, Waxemberg MD, Doñate-Oliver F, Pomata PE, Calvo DJ. Flavonoid modulation of ionic currents mediated by GABA(A) and GABA(C) receptors. *Eur J Pharmacol* 2003;461(2–3):79–87.
- [28] Boissier JR, Dremont C, Robbins R, Pagny J. Tentative de pharmacologie previsionelle dans la domaine des neuroleptiques: actions sédativ centrale et adrenergique de la *N*(dimethoxy-3,4 phenethyl) *N*(chloro-2 phenyl) piperazine. *Arch Int Pharmacodyn Ther* 1961;133:29–32.
- [29] Dunham NW, Miya TS. A note on a simple apparatus for detecting neurological deficit in rats and mice. *J Am Pharm Assoc* 1957;46(3):208–9.
- [30] Lister RG. The use of a plus-maze to measure anxiety in the mouse. *Psychopharmacology* 1987;92:180–5.
- [31] Rodgers RJ, Dalvi A. Anxiety, defence and the elevated plus-maze. *Neurosci Biobehav Rev* 1997;21(6):801–10.
- [32] Pellow S, Chopin P, File SE, Briley M. Validation of open–closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci Methods* 1985;14:149–67.
- [33] File SE, Wardill AG. The reliability of the hole-board apparatus. *Psychopharmacologia* 1975;44(1):47–51.
- [34] Carlini EA, Contar JDP, Silva-Filho AR, Silveira-Filho NG, Frochtengarten ML, Bueno OF. Pharmacology of lemongrass (*Cymbopogon citratus* Stapf). I. Effects of teas prepared from the leaves on the laboratory animals. *J Ethnopharmacol* 1986;17:37–64.
- [35] Randrup A, Munkvad I. Stereotyped activities produced by amphetamine in several animal species and man. *Psychopharmacologia* 1967;11:300–10.

- [36] Swinyard EA, Brown WC, Goodman LS. Comparative assays of antiepileptic drugs in mice and rats. *J Pharmacol Exp Ther* 1952;106(3):319–30.
- [37] Porsolt LD, Anton G, Blavet N, Jalfre M. Behavioral despair in rats: a new model sensitive to antidepressant treatments. *Eur J Pharmacol* 1978;47:379–91.
- [38] Cavadas C, Araujo I, Cotrim MD, Amaral T, Cunha AP, Macedo T. In vitro study on the interaction of *Valeriana officinalis* L. extracts and their amino acids on GABA_A receptor in rat brain. *Arzneimittelforschung* 1995;45:753–5.
- [39] Rodgers RJ, Cole JC. The elevated plus-maze: pharmacology, methodology and ethology. In: Cooper SJ, Hendrie CA, editors. *Ethology and Psychopharmacology*. New York: John Wiley & Sons Ltd.; 1994. p. 9–44.
- [40] Lolli LF, Sato CM, Romanini CV, Villas-Boas LB, Santos CAM, Oliveira RMW. Possible involvement of GABA_A-benzodiazepine receptor in the anxiolytic-like effect induced by *Passiflora actinia* extracts in mice. *J Ethnopharmacol* 2007;111(2):308–14.
- [41] Cryan JF, Kelly PH, Chaperon F, Gentsch C, Mombereau C, Lingenhoehl K, et al. Behavioral characterization of the novel GABA_B receptor-positive modulator GS39783 (*N,N'*-dicyclopentyl-2-methylsulfanyl-5-nitro-pyrimidine-4,6-diamine): anxiolytic-like activity without side effects associated with baclofen or benzodiazepines. *J Pharmacol Exp Ther* 2004;310:952–63.
- [42] Zhao X, Cui XY, Chen BQ, Chu QP, Yao HY, Ku BS, et al. Tetrandrine, a bisbenzylisoquinoline alkaloid from Chinese herb Radix, augmented the hypnotic effect of pentobarbital through serotonergic system. *Eur J Pharmacol* 2004;506(2):101–5.
- [43] Rogawski MA. Mechanism-specific pathways for new antiepileptic drug discovery. In: French J, Leppik I, Dichter MA, editors. *Antiepileptic drug development, advances in neurology*, vol. 76. Philadelphia (PA): Lippincott-Raven Publishers; 1998. p. 11–27.
- [44] Fisas MA, Farré A, Camarasa J, Escubedo E. Effects of lesopitron on the central nervous system arising from its interaction with 5-HT_{1A} receptors. *Pharmacology* 2004;72:57–67.
- [45] Gyertyán I, Sággy K. The selective dopamine D₃ receptor antagonists, SB 277011-A and S 33084 block haloperidol-induced catalepsy in rats. *Eur J Pharmacol* 2007;572(2–3):171–4.
- [46] Haleem DJ, Samad N, Haleem MA. Reversal of haloperidol-induced extrapyramidal symptoms by buspirone: a time-related study. *Behav Pharmacol* 2007;18(2):147–53.
- [47] Neal-Beliveau BS, Joyce JN, Lucki I. Serotonergic involvement in haloperidol-induced catalepsy. *J Pharmacol Exp Ther* 1993;265:207–17.
- [48] Erzín-Waters C, Muller P, Seeman P. Catalepsy induced by morphine or haloperidol: effects of apomorphine and anticholinergic drugs. *Can J Physiol Pharmacol* 1976;54(4):516–9.
- [49] Hauber W, Neuscheler P, Nagel J, Müller CE. Catalepsy induced by a blockade of dopamine D₁ or D₂ receptors was reversed by a concomitant blockade of adenosine A_{2A} receptors in the caudate-putamen of rats. *Eur J Neurosci* 2001;14(8):1287–93.
- [50] Skilbeck KJ, O'Reilly JN, Johnston GA, Hinton T. The effects of antipsychotic drugs on GABA_A receptor binding depend on period of drug treatment and binding site examined. *Schizophr Res* 2007;90(1–3):76–80.
- [51] Ágmo A, Belzung C, Giordano M. Interactions between dopamine and GABA in the control of ambulatory activity. *J Neural Transm* 1996;103(8–9):925–34.
- [52] Keller HH, Schaffner R, Haefely W. Interaction of benzodiazepines with neuroleptics at central dopamine neurons. *Naunyn Schmiedebergs Arch Pharmacol* 1976;294(1):1–7.
- [53] Worms P, Lloyd KG. Biphasic effects of direct, but not indirect, GABA mimetics and antagonists on haloperidol-induced catalepsy. *Naunyn Schmiedebergs Arch Pharmacol* 1980;311(2):179–84.
- [54] Malec D, Langwiński R. The influence of gabaergic system on cataleptogenic action of analgesics and haloperidol. *Pol J Pharmacol Pharm* 1986;38(5–6):501–7.
- [55] Naidu PS, Kulkarni SK. Quercetin, a bioflavonoid, reverses haloperidol-induced catalepsy. *Methods Find Exp Clin Pharmacol* 2004;26(5):323.
- [56] Santos KC, Santos CAM, Oliveira RMW. *Passiflora actinia* Hooker extracts and fractions induce catalepsy in mice. *J Ethnopharmacol* 2005;100(3):306–9.
- [57] Quéva C, Bremner-Danielsen M, Edlund A, Ekstrand J, Elg S, Erickson S, et al. Effects of GABA agonists on body temperature regulation in GABA_B(1)–/– mice. *Br J Pharmacol* 2003;140:315–22.
- [58] Takeda H, Tsuji M, Matsumiya T. Changes in head-dipping behavior in the hole-board test reflect the anxiogenic and/or anxiolytic state in mice. *Eur J Pharmacol* 1998;350(1):21–9.
- [59] Rego JC, Viana AF, Le Maître E, Deniel A, Rates SMK, Leroux-Nicollet I, et al. Comparisons between anxiety tests for selection of anxious and non anxious mice. *Behav Brain Res* 2006;169(2):282–8.
- [60] Andrews N, File SE. Handling history of rats modifies behavioural effects of drugs in the elevated plus-maze test of anxiety. *Eur J Pharmacol* 1993;235(1):109–12.
- [61] File SE. Tolerance to the behavioral actions of benzodiazepines. *Neurosci Biobehav Rev* 1985;9:113–22.
- [62] Calil CM, Marcondes FK. The comparison of immobility time in experimental rat swimming models. *Life Sci* 2006;79(18):1712–9.
- [63] Flugy A, Gagliano M, Cannizzaro C, Novara V, Cannizzaro G. Antidepressant and anxiolytic effects of alprazolam versus the conventional antidepressant desipramine and the anxiolytic diazepam in the forced swim test in rats. *Eur J Pharmacol* 1992;214(2–3):233–8.
- [64] Nakazawa T, Yasuda T, Ueda J, Ohsawa K. Antidepressant-like effects of apigenin and 2,4,5-trimethoxycinnamic acid from *Perilla frutescens* in the forced swimming test. *Biol Pharm Bull* 2003;26(4):474.
- [65] Clenet F, Hascoet M, Fillion G, Galons H, Bourin M. Role of GABA-ergic and serotonergic systems in the anxiolytic-like mechanism of action of a 5-HT-moduline antagonist in the mouse elevated plus maze. *Behav Brain Res* 2005;158(2):339–48.
- [66] Starr KR, Price GW, Watson JM, Atkinson PJ, Arban R, Melotto S, et al. SB-649915-B, a novel 5-HT_{1A/B} autoreceptor antagonist and serotonin reuptake inhibitor, is anxiolytic and displays fast onset activity in the rat high light social interaction test. *Neuropsychopharmacology* 2007;32:2163–72.
- [67] Manach C, Donovan JL. Pharmacokinetics of dietary flavonoids in humans. *Free Radic Res* 2004;38(8):771–85.
- [68] Shimoi K, Okada H, Furugori M, Goda T, Takase S, Suzuki M, et al. Intestinal absorption of luteolin and luteolin 7-O-β-glucoside in rats and humans. *FEBS Lett* 1998;438:220–4.