

A Review of Autism and the Immune Response

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The autistic spectrum disorders (ASD) are complex developmental disorders that manifest during childhood. By definition, children with ASD are characterized by qualitative impairments in social interaction, deficits in verbal and non-verbal communication, and with restricted repetitive and stereotyped patterns of behavior and interests (American Psychiatric Association, 1994). ASD describes a range of conditions including autism and Asperger's syndrome that fall within the umbrella term of pervasive developmental disorders (PDD). The prevalence of ASD has increased substantially over the last decade rising from 5 to 60/10,000 (Fombonne, 1999; Bertrand et al., 2001; Chakrabarti and Fombonne, 2001; Fombonne et al., 2001; Wing and Potter, 2002). The male to female ratio in autism are also skewed with a three- to four-fold increased incidence in boys. In the majority of cases the cause of ASD are unknown, although strong genetic links have been shown for cases with tuberous sclerosis, neurofibromatosis, chromosomal abnormalities and fragile X. In addition, environmental factors have been implicated for prenatal rubella infection, anticonvulsants and antiemetics taken during pregnancy, perinatal hypoxia, post-natal infections such as encephalitis and in metabolic disorders such as phenylketonuria (Fombonne et al., 2001; Baird et al., 2003). Evidence for the contributing role of genetic factors in ASD have also come from monozygotic twin and family studies, with concordance rates for broad spectrum ASD of 90% and a familial risk of 5-10 times higher than the general population, respectively (Bailey et al., 1995; Szatmari, 1999; Rutter, 2000). However, within the general population there appears to be no bias or selective pressure against ASD traits, where arguably from the evolutionary viewpoint due to impairments in social interactions and in some cases lifelong dependency, one would anticipate selective pressure would exist. As such, it has been suggested through genomic screens, that in autism there are multiple weak gene interactions. Indeed, many candidate genes that determine disease susceptibility, or severity, or disease course, have been

implicated and studied in autism. There exists a great variability in gene expression (Szatmari, 1999; Risch et al., 1999; Rutter, 2000), with the most likely genetic models predicting multiple gene interactions of up to 15 or more with low individual contributions. However, unequivocal evidence for a genetic association with ASD has so far not been established. Moreover, with the increased prevalence of autism the influence of a definitive set of genetic determinants has become more difficult to determine. The lack of association between ASD and social class, immigrant status and geographical location may suggest that a biological i.e. not solely genetic cause, plays a leading role in the etiology of autism. Furthermore, features such as epilepsy, clear clinical regression of development, mental retardation, persistent neurological signs such as primitive reflexes (Volkmar et al., 2003) and gastrointestinal symptoms occur in some but not in all cases. Thus there may exist subsets of ASD patients, with separate multiple etiologies that affect biological and psychological functioning, that manifest with differing clinical phenotypes.

There is a growing awareness of an immunological involvement in children with ASD. Systemic immunologic aberrations in ASD have been linked with both autoimmunity, describing antibodies reactive for CNS proteins with the potential for neuronal tissue destruction, and secondly, with dysfunctional immunity such as abnormalities or deficits of function in immune cell subsets, leading to an inappropriate or ineffective immune response to pathogen challenge. Indeed, in general the links between the immune and neurological systems are becoming increasingly well known. Products of the immune system may play an important role in early neurodevelopment and can influence patterns of behavior. The immune and nervous system are both complex, highly evolved systems preserved within structural networks that transfer signals through the release of chemical mediators such as neuropeptides and cytokines. In addition, the neuronal synapse and the contact interface between T cells and antigen presenting cells share a structurally similar

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architecture. There is continual cross communication between the immune and nervous systems, with many peptides playing a role in both. In ASD, through the analysis of neuropeptide and neurotransmitter levels, many neuroactive compounds that also share immunomodulatory properties, have been implicated in the disease process.

NEUROTRANSMITTERS AND NEUROIMMUNOLOGY

Neuropathologic findings in autism include reduced numbers of neurons and reduced dendritic arborisation in the amygdala, hippocampus, mammillary bodies and medial septial nucleus (Kemper and Bauman, 2002). In addition, observations of increased brain size compared with controls and decreased numbers of Purkinje and granule cells have also been noted in autism (Volkmar et al., 2003). These findings clearly indicate that there are neurological involvements in ASD that affect the development and differentiation of neurons in the brain. It has been proposed that increased or abnormal levels of neuroactive compounds early during neurodevelopment may lead to autism. Indeed, abnormalities in various neurotransmitter systems have been implicated in the development of autism, including serotonin, dopamine, noradrenaline (norepinephrine), gamma-aminobutryic acid (GABA), glutamate and neuropeptides. Such peptides as oxytocin and vasopressin which are involved in immune tolerance, serotonin (5-HT) which exerts concentration dependent suppressive or proliferative responses on T cells, and the immunomodulating vasoactive intestinal peptide (VIP), have all been implicated in ASD (Nelson et al., 2001; Sharp, 2003).

Serotonin is a biogenic amine that has wideranging effects on numerous physiological processes such as circadian rhythyms, appetite, mood, sleep, anxiety, motor activity and cognition. Serotonin is found in serotonergic neurons of the nervous system, enterochromaffin and neuroendocrine cells of the endocrine system as well as in platelets and on lymphocytes of the immune system. Consistent findings of increased peripheral blood platelet serotonin levels have been demonstrated in approximately one third of children with autism (Anderson et al., 1990; Singh et al., 1997), while selective 5-HT reuptake inhibitors have been shown to be beneficial with regard to treating obsessional and repetitive behavioral phenotypes in some patients (McDougle et al., 1996). The mechanism by which platelet 5-HT is elevated remains unknown, but it is likely that the effect is due to alterations in the platelets themselves resulting in increased platelet uptake or reduced platelet 5-HT2 receptor binding (Cook and Leventhal, 1996). Interestingly, auto-antibodies directed toward serotonin receptors have been found in autistic patients compared with controls (Todd and Ciaranello, 1985; Singh et al., 1997), suggesting an abnormal immune response may be involved in serotonin regulation. In addition, enzymes that control the conversion of tryptophan into serotonin or the formation of kynurenine derivatives are under the control of cytokines such as IFNy and IL-1 (Dunn et al., 1999; Wirleitner et al., 2003). Typtophan depletion has previously been shown to result in an increase of autistic behaviors (McDougle et al., 1996). Thus, accelerated tryptophan degradation as a result of immune activation could lead to an increase in autistic features. In autistic boys, decreased measurements of serotonin synthesis in the frontal cortex and thalamus, but elevated synthesis in the contralateral dentate nucleus were observed using PET scans (Chugani et al., 1999). However, factors such as the age or gender of the subject and associated mental retardation may alter serotonin levels. Nonetheless it has been proposed that there exists a normal developmental process characterized by high serotonin levels in typically developing children up to 5 years of age, which is disrupted in autistic children (Chugani et al., 1999). Encouraging preliminary data have suggested a link between the 5-HT transporter gene and autism, but so far the nature of the specific link remains elusive (Anderson et al., 2002; Conroy et al., 2004; Nabi et al., 2004). Similarly, changes in blood levels of norephinephrine, aspartic acid, glutamine, glutamic acid and GABA have been shown in individuals with autism compared with controls (Cook et al., 1990; Rolf et al., 1993), however, further large scale studies with patients matched for age and behavioral characteristics need to be performed.

Opioid peptides and opioid receptors are important modulators of neural development, influencing migration, proliferation and differentiation within the CNS (Zagon, 1987). It has been hypothesized that an excess of opioid peptides will have detrimental effects on brain development and behavior, and that autism may be a result of abnormal levels or activity of opioid peptides. Not only are immune cells capable of producing opioid peptides, but they also have opioid receptors on their cell surface. The precise mechanisms that underlie the immunosuppressive effects of opioids remain unknown, however, it has been proposed that they can operate as cytokines (Peterson et al., 1998), acting through receptors on peripheral blood and/or glial cells. For example, the anti-inflammatory kappa-opioid drugs are able to reduce adhesion molecule expression, inhibit cellular trafficking, reduce TNF-a release and expression, and alter mRNA expression of neuropeptides, presumably through opioid-receptor interactions involved in different phases of an immune response (Walker, 2003). In contrast, beta-endorphin increases the expression of complement and Fcy receptors on the surface of neutrophils and induces concentration dependent stimulation of reactive oxygen intermediates (Menzebach et al., 2003). The actions of beta-endorphin on neutrophils are mediated through the N-terminal fragment, and not the C-terminal fragment. Studies on beta-endorphin in autism have found increased C-terminal and reduced N-terminal beta-endorphin fragments (Leboyer et al., 1994). However, a clear pattern of beta

endorphin levels has yet to be established as further studies have found increased, decreased and normal levels in plasma and CSF (Croonenburghs et al., 2002). Another important source of opioids with neurmodulating activity is from the diet. Beta-casemorphine-7, an opioid exclusively of dietary origin, has been shown to be present in patients with pyschoses including autism (Wakefield et al., 2002). Indeed, the beneficial effects on autistic behavior following dietary exclusion therapy are thought, in part, to be a result of reduced opioid intake (Knivsberg et al., 1995; Whitely et al., 1999; Knivsberg et al., 2002). Furthermore, therapeutic trials using the oral opioid antagonist, naltrexone, in autism have shown improvements in behavioral characteristics such as repetitive stereotypies, social contact, and self-injury (Herman et al., 1987; Panksepp et al., 1991; Zingarelli et al., 1992). How the immune system is affected by these therapeutic interventions is largely still unknown.

Neuropeptides are small, biologically active peptides derived from the central and peripheral nervous system. Through animal models, it has been suggested that the neuropeptides oxytocin and vasopressin play a critical role in the regulation of social recognition, affiliation and attachment (Young et al., 2002). Prairie voles are social animals who form long lasting pair bonds. Central infusion of oxytocin in females or vasopressin in males helps to establish partner bonding, a phenomenon that can be blocked using specific antagonists (Williams et al., 1994; Young et al., 2001). In prairie voles, oxytocin and vasopressin receptors are located in the ventral forebrain whereas, in contrast, asocial or solitary montane voles which do not form bond pairs have different patterns of expression for oxytocin receptors (Young et al., 2002). Therefore, the pattern of receptor distribution together with the concentration of released neuropeptides are important in the establishment of socially rewarding interactions. Furthermore, oxytocin knockout mice have normal cognitive abilities but have diminished social recognition (Ferguson et al., 2000). In autism, plasma levels of oxytocin are reported to be significantly lower than controls (Modahl et al., 1998). In addition, further analysis of neuropeptides in neonatal blood by recycling immunoaffinity chromatography found that concentrations of VIP, calcitonin-gene related peptide, brainderived neurrotrophic factor and neurotrophin 4/5 were increased in ASD compared with typically developing control children but could not be distinguished from those with mental retardation (Nelson et al., 2001). So far, signature patterns of neurpeptides and neurotransmitters in autism have yet to be established.

It is increasingly clear that besides their function as neurotransmitters, neuropeptides have an influence on almost all body functions including the immune system. For example, oxytocin is present in the thymus at high levels and is expressed on thymic epithelial cells, where it is thought to act both as a self-antigen and as a promoter of T cell focal adhesion (Gimpi and Fahrenholz, 2001) and as such plays a role in immune tolerance. In addition,

Dunzendorfer et al. have suggested a role for neuropeptides as immunomodulators and have presented a novel concept of neuropeptide-mediated regulation of dendritic cell (DC) migration. They investigated locomotion of mononuclear cell-derived DCs at different maturation stages toward gradients of sensory neuropeptides in vitro. Calcitonin gene-related peptide, VIP, secretin, and secretoneurin induced immature DC chemotaxis comparable to the potency of the chemokine RANTES, whereas substance P and macrophage-inflammatory protein-3 beta stimulated immature cell migration only slightly. Moreover, there was a distinct neuropeptideinduced signal transduction seen in DC exposed to neuropeptides (Dunzendorfer et al., 2000). In addition, it has been shown that VIP synergizes with TNF- α in the induction of DC maturation (Delneste et al., 1999). The notion that a defect in DC chemotaxis could have ramifications on the normal process of immature DC migration to sensory nerve fibers where they participate in inflammatory events is intriguing in the context of autism. In the central nervous system (CNS) DCs have been found in normal meninges, the choroid plexus, and cerebrospinal fluid. During inflammation in the CNS, there is a recruitment of DCs. It has been speculated that they may equally play a role in the defense against infections and contribute to the break-down of tolerance to CNS autoantigens (Pashenkov et al., 2003). Microglia cells, another CNS cell lineage, are antigen presenting cells (APCs) that share similarities with both DCs and macrophages. Microglia cells, as part of the CNS innate immune system, participate in many reactive processes in the CNS and have been implicated in the exacerbation of neurological conditions such as multiple sclerosis, Alzheimer's disease, AIDS, and viral encephalitis (Nelson et al., 2002). In the autoimmune encephalomyelitis (EAE) model and in patients with multiple sclerosis, self-myelin antigens were observed in cells expressing DC/macrophage markers, MHC class II, and co-stimulatory molecules, a finding not present in controls (de Vos et al., 2002). Taken together, these findings suggest a central role for DC and neuropeptide mediated chemotaxis in the control of CNS inflammation and modulation of T cell reactivity against CNS antigens.

Cytokines and other products of immune activation have widespread effects on neuronal pathways, and in part may be responsible for many common features of ASD such as mood and sleep disturbances. The brain is protected and isolated by the blood brain barrier from potentially deleterious agents present in the blood including inflammatory immune cells and proteins. However, recent data have suggested that the brain is an immunologically active organ and that immune reactions do occur within the CNS. The entry of cytokines produced in the periphery into the brain is predominantly restricted by the blood brain barrier. However, they may gain access through active transport mechanisms or at circumventricular regions where the barrier is less controlling (Wilson *et al.*, 2002). In addition, cytokines may affect

brain barrier function directly by binding to receptors on brain endothelial cells leading to the impairment of function (Wilson et al., 2002). Evidence has emerged that leuckocytes migrate into the brain from the blood via the cerebral spinal fluid to the choroids plexus, or from the blood to either the subarachnoid space or parenchymal perivascular space (Ransohoff et al., 2003). These leuckocytes may then act as a potential source of cytokines. Alternatively, afferent neurons may be directly responsive to peripheral cytokine stimulation (Dantzer et al., 1998). Indeed, cytokines can affect enteric glial cells (Von Boyen et al., 2004). The effects of cytokines on the mood, sleep, nutritional intake, exploratory behavior, and social interactions do not necessarily have to result in neuronal damage or death. Systemic cytokines administered at therapeutic doses, such as IFN-α, IL-2 and TNF- α , have side effects including mood depression, sleep disorder, impaired cognitive function, decreased exploratory behavior, and changes in motivation (Licinio et al., 1998; Larson, 2002). Systemic cytokine administration can also cause increases in noradrenergic, dopaminergic and serotonergic metabolism in the hypothalamus, hippocampus and nucleus accumbens (Mohankumar et al., 1991; Shintani et al., 1993; Merali et al., 1997). Cytokines can activate and exert trophic effects on glial cells, and in turn, glial cells can produce cytokines and chemokines upon activation. As the CNS is largely populated by astroglia and microglial cells, these cytokine-cell interactions have important implications for neuronal cell functioning and development.

SYSTEMIC IMMUNE ABNORMALITIES IN ASD PATIENTS

Various immune abnormalities have been described by an increasing number of laboratories worldwide with findings consistently showing both the involvement of autoimmune processes and the dysregulation of the immune response in children with autism. Many autistic children suffer prolonged and recurrent infections, as well as reduced responsiveness to common recall antigens on cutaneous delayed hypersensitivity tests compared with controls (Murch et al., 1999). Immune aberrations consistent with a dysregulated immune response reported in ASD children include decreased peripheral lymphocyte numbers (Ashwood et al., 2003), decreased response to T cell mitogens (Stubbs and Crawford, 1977; Warren et al., 1986), incomplete or partial T cell activation evinced by increased numbers of DR+T cells without the expression of the IL-2 receptor (Plioplys et al., 1994; Denney et al., 1996), dysregulated apoptosis mechanisms (Engstrom et al., 2003), and the imbalance of serum immunoglobulin levels including increased IgE and decreased serum IgA (Ashwood et al., 2003). In initial cytokine analysis studies of 10 autistic children Stubbs (Stubbs, 1995) showed increased levels of plasma IFN-α compared with 4 adult controls. In a further study, plasma levels of IL-12 and IFNy were increased in autistic children but there were no differences between IL-6, TNF α and IFN α when compared with controls (Singh, 1996). These studies proposed that there are aberrant T cell cytokines profiles in children with ASD compared with controls. However, there is an apparent divergence of opinion on the predominant polarity of the dysregulated immune response. In cell culture studies, Gupta showed intracellular cytokine levels of stimulated IL-2 and IFNy expressing CD4+ and CD8+ cells to be lower in 20 autistic patients compared with 20 aged matched controls. In contrast, the number of CD4+ and CD8+ cells expressing IL-4 were increased (Gupta et al., 1998). Croonenburghs et al. (2002) reported increased IFNy, IL-6 and IL-1ra in unstimulated culture supernatants from ASD patients, although no information on experimental design or details of autistic patients or controls were given. More recently, plasma IFNy levels in 29 autistic children correlated positively with the generation of the potent functional and developmental intercellular CNS messenger nitric oxide (NO), a finding indicative of NO-related oxidative stress (Sweeten et al., 2004). A similar mechanism of IFNy induced plasma neopterin, may be responsible for the elevated levels observed in autistic patients and may reflect cell-mediated immune activation (Sweeten et al., 2003). Evidence for an increased inappropriate innate immune response was shown by the over-production of pro-inflammatory cytokines, in particular TNFα, in peripheral blood mononuclear cells stimulated with bacterial LPS in ASD children compared with controls (Jyonouchi et al., 2001). Interestingly, siblings also showed raised TNFα levels compared with controls suggesting a genetic defect in control of innate immune responses in children with ASD and their family members. Furthermore, decreased NK cell activity has been shown in ASD children and is further indicative of a functional deficit in the innate immune response (Warren et al., 1987). However, very few studies on the innate system in patients with autism have been performed thus far. Other findings in autistic subjects, with an immunological significance, include increased frequency of the null alleles of complement and HLA-DRB1*04 allele and low levels of C4b complement protein (Warren et al., 1991, 1994, 1996). However, it is hard to ascertain a clear signature immune cellular or cytokine response in the pathology of early childhood development seen in autism. So far there are a lack of systematic studies with sufficiently large enough sample sizes. In addition, many studies are not adequately matched for the age and gender of subjects and controls, where autistic children are often compared with much older controls. In part, these difficulties occur as there is often limited access to autistic populations and the behavioral aspects of autism make sampling difficult. Furthermore, the young age of the subjects makes rigorous and wide-ranging immunological studies prohibitive. A further consideration is that it is now becoming increasingly apparent that autistic patients may be categorized into various phenotypes based

on differing patterns of genetic, immunologic and behavioral aspects. In the future, well defined studies with fully characterized patient and matched control groups will need to be performed in order to ascertain how the immune response is involved in autistic pathology.

AUTOIMMUNE PROCESS IN ASD

The presence of antibodies directed against components of the CNS in the sera of autistic children, is indicative that a possible autoimmune process may be involved in the pathology of ASD. Autoimmune diseases arise when the immune system is inappropriately directed to recognize and exert an exaggerated respond to "self" components. These include, but are not restricted to diseases such as; myasthenia gravis, multiple sclerosis, systemic lupus erythematosus, primary biliary cirrhosis, and Graves thryrotoxicosis. The exact mechanism of autoimmunity in these diseases is not identical, but they all possess autoreactive antibodies and T cells. Various anti-brain antibodies have been found in autistic patients including autoantibodies to serotonin receptor (Todd and Ciaranello, 1985), myelin basic protein (MBP) (Singh et al., 1993), neuron axon filament protein (NAFP) (Singh et al., 1997), cerebellar neurofilaments (Plioplys et al., 1994), nerve growth factor (Kozlovskaia et al., 2000), alpha-2-adrenergic binding sites (Cook et al., 1993), antibrain endothelial cell proteins (Todd et al., 1988; Connolly et al., 1999) and antibodies against the caudate nucleus (Singh and Rivas, 2004). In a recent paper, serum from a mother with an autistic child found to bind to Purkinje cells and other neurons, when injected into gestating mice, induced behavioral changes including altered exploration, motor coordination and changes in cerebrallar magnetic resonance spectroscopy in the offspring. In contrast, mice injected with sera from mothers with typically developing children showed no behavioral changes (Dalton et al., 2003). This study supports the notion that maternal antibodies may influence the neurodevelopmental process in autism. In addition, the frequency of autoimmune disorders in family members of 61 ASD children and 46 typically developing normal controls were compared (Comi et al., 1999). The frequency of autoimmune disorders was increased in the ASD group with over 40% of families having two or more close family members with autoimmunity. Findings that have since been replicated by Sweeten et al. (2003) in families of pervasive developmental disorder probands. However, these studies were based on self-reporting rather than medical records and thus may have a significant margin of reporting error.

The pathophysiological significance of auto antibodies reported in children with autism is uncertain. For example increased autoantibodies would suggest that there is increased neuronal damage, as is the case in multiple sclerosis where, following demyelination, MBP is unmasked and there is a subsequent generation of antibodies. To date however, evidence of demyelination in autism has remained elusive (Rumsey and Ernst, 2000). Glial fibrillary acidic protein (GFAP) measured in the CSF of 47 autistic children was significantly elevated compared to age matched 10 control children, suggesting that gliosis and unspecific brain damage may occur in autism (Ahlsen et al., 1993). However, as GFAP correlates strongly with age, due most likely to age dependent expansion of fibrillary astrocytes, caution must be shown in interpreting these data (Rosengren et al., 1992). Another possible cause for the development of autoimmunity is a dysfunction in immune regulation, that allows immune responses to continue unchecked. One regulatory T cell population is identified by the combination of the CD4 (T helper cells) and CD25 (IL-2 receptor) cell surface markers, which are a cell population that as discussed above, is at a low frequency in autism. A dysregulation of IL-10 and TGFB1 release parallel to times when pro-inflammatory signals such as TNF α are high, will lead to an imbalance of inflammatory signals. Furthermore, defects in apoptosis mechanisms will promote the survival of immune cells including autoreactive lymphocytes. Dysregulated FAS mediated cell death and increased soluble FAS receptor have been described in autism (Engstrom et al., 2003). Observations of elevated anti-CNS antibodies in autism, are at best unconfirmed, and in some cases, for example serotonin receptors and MBP, markedly conflicting. However, taken together the findings of autoimmunity in families and the plethora of anti-brain antibodies suggest that in some patients, autoantibodies that target the CNS may be a pathological or exacerbating factor in neuronal development in children with ASD. It is also important to note that for each antibody tested the number of autistic children showing positivity was far from 100%. It is also unclear whether individual autistic children are positive for more than one antibody. Again, it can be inferred that increased autoimmunity may be confined to a subset of autistic patients. Indeed, large cohort studies with thoroughly defined and specifically phenotyped autistic patient groups and well matched age and sex controls, need to be performed to confirm the potential role of autoantibodies in the pathology of either all or subsets of autistic patients. Moreover, the development of an animal model will be crucial to determine the role of autoantibodies in the pathology of autism.

RESPONSE TO VIRUSES

Many infectious agents including; rubella, measles, human herpesvirus 6 (HHV-6), influenza and cytomegalovirus have been associated with the aetiology of autism (Chess *et al.*, 1978; Binstock, 2001; Yamashita *et al.*, 2003). These associations, although not conclusive for all children with ASD, may highlight an underlying inability of ASD children to fully eradicate viral insults. As such,

viruses and viral products may persist in children with ASD. Indeed, defects in T cell numbers and function, a phenomenon observed in ASD, may compromise the body's defense system to infectious agents and engender the body susceptible to recurrent infections. Persistent viral infection has been described in mucosal tissue of ASD children compared with age-matched typically developing controls, with viral RNA present in DC populations (Uhlmann et al., 2002). This persistence may be indicative of a primary failure to successfully eradicate antigen and the inability to deal with viral insults. Furthermore, increases in lymphoid tissue seen at mucosal sites in ASD children may be reflective of a local inflammatory response to the presence of a persisting Ag (Wakefield et al., 2000; Ashwood et al., 2003). Paramyxoviruses, such as measles have profound effects on DC function including the inhibition of LPS stimulated IL-12 release. In addition, the ability to stimulate T cell proliferation by mitogen, was greatly reduced in the presence of measles infected DCs (Klagge and Scheider-Schaulies, 1999). The migration of infected DCs to T cell areas of lymphoid organs could therefore exert immunosuppressive effects on the surrounding T cells.

In a mouse model, viral infections in pregnant mothers leave long-lasting effects on the offspring, including an increased risk of behavioral abnormalities with particular relevance to autism and schizophrenia (Patterson, 2002). In pregnant mice (BALB/c and C57BL/6 strains), when human influenza virus was administered, offspring displayed aberrant behavioral responses as adults, with deficits in prepulse inhibition (PPI) in the acoustic startle response, findings also observed in autistic subjects (Fatemi et al., 2002). It is hypothesized that maternal immune responses generated to eradicate the viral insult affect fetal brain development and the upregulation of specific cytokines are implicated at this time. In another animal model, infection with Borna disease virus (BDV) in Lewis rats leads to behavioral and CNS abnormalities reticent of neurodevelopmental disorders such as schizophrenia and autism in humans (Hornig et al., 1999). Borna disease virus infection primarily targets the hippocampal neurons, affecting ERK 1/2 phosphorylation, the expression of synaptic vesicle proteins, and hampers synaptogenesis and synaptic organization induced by the neurotrophin brain-derived neurotrophic factor (BDNF). However, BDNF receptor expression, neuronal survival and morphology are not affected (Hans et al., 2004). These findings show that BDV acts in a non-cytopathic way to alter neuronal growth factor mediated events and thereby alter synaptic connections between neurons, possibly by interfering with cell signaling events downstream of receptor binding, where such alterations may affect all functions of the brain. These models of infection will provide valuable insight into the mechanisms of human neurodevelopmental disorders and the interplay of the developing neuronal and immune system in the pathology of environmentally induced behavioral abnormalities.

Bacterial and viral infections have been suspected in the triggering of or excacerbation of autoimmune diseases (Panoutsakopoulou and Cantor, 2001). In autism, patients with anti-measles IgG antibodies were also positive for anti-MBP (90%) and anti-NAFP (73%) antibodies, a similar relationship were seen in patients with positive titers for anti-HHV-6 antibodies and corresponding anti MBP and anti-NAFP antibodies (Singh et al., 1998). Interestingly, in these studies anti-measles and anti-HHV-6 antibodies were not different between autistic patients and controls. The underlying mechanism between viral insults and disease in autism remains unclear. Pathogenesis may result due to cellular damage induced as a result of immune responses aimed at eradicating an invading virus. Equally, impaired immune responses could increase susceptibility to infections in autistic children, or an inability to clear viruses leading to persistent infections that could then subvert immune function or act directly upon the CNS to elicit neurodevelopmental abnormalities. The study of pathogen-induced immune responses and the pattern of pre- and post-natal exposures on the developing neuronal system may provide interesting data.

MUCOSAL IMMUNITY IN ASD PATIENTS

Gastrointestinal symptoms have been described in a number of ASD patients, in whom symptoms include abdominal pain, bloating, diarrhoea and constipation (Horvath et al., 1999; Afzal et al., 2003). The precise number of children with ASD who have gastrointestinal symptoms is not currently known, but some studies have estimated this to be between 18 and 40% (Baron-Cohen et al. 1996; 2000; Horvath et al., 1999; Fombonne et al., 2001). Reported gastrointestinal abnormalities include low activities of disaccharidase enzymes, defective sulphation of ingested phenolic amines (paracetamol), bacterial overgrowth with greater diversity and number of clostridial species, more numerous Paneth cells, increased intestinal permeability and a positive effects on behavioral cognition following dietary intervention (Knivsberg et al., 1995; 2002; D'Eufemia et al., 1996; Alberti et al., 1999; Horvath et al., 1999; Finegold et al., 2002). Clinical and pathological studies have described an apparently characteristic gastrointestinal immunopathology in this subset of ASD children (Wakefield et al., 1998, 2000; Furlano et al., 2001; Torrente et al., 2002), in which chronic ileo-colonic lymphoid nodular hyperplasia (LNH) and entero-colitis are key features. The mucosal lesion consists of a pan-enteric lymphocytic infiltrate, with a variable degree of acute inflammation and eosinophil infiltration (Wakefield et al., 1998, 2000; Furlano et al., 2001; Torrente et al., 2002). Flow cytometric and immunohistochemical analyses of mucosal lymphocyte populations in ASD children have demonstrated qualitatively consistent abnormalities at different anatomical sites including stomach, duodenum, ileum and colon

(Furlano et al., 2001; Torrente et al., 2002; Ashwood et al., 2003), indicating a relatively homogenous mucosal lymphocyte infiltrate. Although LNH is not an uncommon finding in children with allergies or immunodeficiency, there is increased frequency and severity in patients with autism (Wakefield et al., 2002). In addition, findings of focal deposition of serum IgG from ASD children which co-localises with complement C1q on the basolateral enterocyte membrane, changes not seen in histologically normal and inflamed mucosa of developmentally normal children or children with cerebral palsy (Torrente et al., 2002), and increased intestinal permeability in ASD (D'Eufemia et al., 1996) is suggestive of an inflammatory process that may perturb the intestinal barrier function in ASD children. Co-localisation of immunoglobulin and complement components on the epithelial membrane have been found in both stomach and duodenal specimens and may be indicative of an autoimmune process directed against self-antigen contained within epithelial cells (Torrente et al., 2002). Furthermore, increased basement membrane thickness and abnormal patterns of epithelial glycosaminoglycans have been seen in autistic children compared with controls and may be indicative of inflammatory degradation that could contribute to disruption of the intestinal barrier function (Furlano et al., 2001).

While there is a great deal of speculation, the exact mechanism of how mucosal changes may influence autistic development or behavior is still not clear. It may be that antigens in the diet can cross the mucosa more easily through the disrupted intestinal barrier, where they cause local inflammatory reactions generating proinflammatory cytokine signals that interact with afferent neurons. Another possibility is that the failure to detoxify neuroactive antigens from the gut may lead to cognitive impairments. Similarly, increased passage of exorphins and/or opioids from the diet such as gliadomorphin and casomorphin into the body, where they can interact directly with the CNS, may play a role in inducing behavioral features of autism. In a further theory, reduced ileal absorption of vitamin B₁₂, a necessary cofactor for nerve myelogenesis and the formation of myelin (Wakefield et al., 1998; Cornish, 2002), could lead to impaired nerve function, which again highlights how altered mucosal integrity may induce neuronal and behavioral changes. However, conclusive evidence to answer how disrupted intestinal mucosal functions affect neurodevelopment and the CNS is still necessary.

In peripheral blood, elevated pro-inflammatory cytokines have been reported to be produced upon stimulation with dietary proteins in autistic children compared with controls (Jyonouchi *et al.*, 2001). In some studies, circulating antibodies to food substances, namely casein and gliadin have been found (Reichelt *et al.*, 1990; Lucarelli *et al.*, 1995). However, these antibodies are also found with similar frequency in typically developing population. Furthermore, antibodies to neuronal specific antigens in the sera of children with autism could cross

react with dietary peptides including milk butyrophilin, streptococcus M protein and chlamydia pneumoniae (Vojdani et al., 2002), suggesting that bacterial infections and milk antigens may modulate the autoimmune process in autism. Interestingly, mucosal lymphocytes isolated from the duodenum, ileum and colon of ASD patients showed increased spontaneous production of pro-inflammatory intracellular cytokines, most notably TNFα, when compared with aged matched controls, including those with similar symptoms of constipation (Ashwood et al., 2002, 2003). In addition, of particular interest was the inverse relationship between raised numbers of pro-inflammatory TNF α positive lymphocytes and decreased numbers of cells expressing the regulatory cytokine IL-10 in ASD children. These findings support the hypothesis that there is mucosal immune dysregulation with a pro-inflammatory lymphocyte cytokine profile, in ASD children. However, in a study that included one autistic child and two with ASD, macrophage release of monokines including IL-1β from ex vivo biopsy explant cultures into culture supernatant was not statistically different when compared with nine older children (Defelice et al., 2003). These data may indicate differential roles for activated lymphocytes and antigen presenting cells in the entero-colitis associated with some autistic children. In an open-label trial, administration of vancomycin, an antibiotic that is poorly absorbed, resulted in objective cognitive improvements in autistic children (Sandler et al., 2000), presumably due to treatment of intestinal dysbiosis. However, once administration of vancomycin was ceased, the patients cognitive functions regressed suggesting that the initial improvement was through beneficial effects on the intestinal pathology (Wakefield et al., 2002). The exacerbation of gastrointestinal and behavioral symptoms in autism induced by certain foods, particularly those containing gluten and casein, has been shown through dietary intervention and their removal from the diet (Knivsberg et al., 1995, 2002). Autistic children on gluten and casein free diets also showed significantly lower eosinophil infiltrate in intestinal biopsies compared with those on a conventional diet (Ashwood et al., 2003). The significance of this finding is still unclear. However, it has been recently proposed that immune responses associated with allergy may contribute to the pathogenesis of autoimmune diseases of the CNS in both humans and in animal models (Pedotti *et al.*, 2003).

IMMUNE THERAPY

Various studies have demonstrated that immunotherapy may be effective in the treatment of some children with autism (Gupta *et al.*, 1996; Plioplys, 1998; Gupta, 2000; Wank, 2002). As stated above, studies that have shown cognitive benefit from dietary intervention have been based upon the assumed removal of substrates for certain neuroactive opioid peptides present in the food

(Knivsberg et al., 1995, 2002). Alternatively, although not exclusively, food antigens may provide a pro-inflammatory stimulus in some ASD children, as suggested by the data of Jyonouchi et al. (2001). Additionally, alterations in gut flora induced by dietary changes or by the administration of antibiotics may lead to beneficial effects. Further studies on immune based therapies including the administration of transfer factor, treatment with pentoxifylline, and intravenous immune globulin treatment have shown improvements in some autistic children (Gupta, 2000). However, these studies remain to be confirmed in larger placebo controlled, double-blinded, crossover studies with well matched control and subject groups. Furthermore, in depth behavioral assessments before and after treatment need to be performed.

CONCLUSIONS

While there is a growing cognizance of immunological dysfunction in some ASD children, a great deal of research remains to be done. There are several aspects of the immune phenomenology thus far reported in ASD that require further investigation. Among these is the suggestion that patients with ASD have an altered or inappropriate response to viruses, the nature, extent and involvement of autoimmune processes in disease, and the importance of mucosal inflammation co-morbidity in some children. Each of these potentially pathological processes may be involved in the aetiology of autism in only a subset of children, or to varying degrees in all. Among the current and highly controversial data, there is much speculation, but little is truly known about the response of children with ASD to foreign antigens. The link between the immune and neurological systems is becoming increasingly well known. Products of the immune system play an important role in early neurodevelopment and can influence patterns of behavior. Bidirectional communication between the immune and nervous systems with many neuropeptides and neurotransmitters playing a role in both, is also of great interest. How the various immune abnormalities that have been reported in patients with ASD, are related to the development of neurologic changes is not yet known. In addition, exposure of the developing neuronal system to enhanced or aberrant immune responses during critical periods of development may not be uniform and may lead to the expression of different phenotypes in autistic patients, dependent upon rates of neuronal and immune development and genetic composition. So far, no clear signature patterns of cellular or cytokine responses have been found in autistic patients that can identify autism from other neurodevelopmental disorders, nor identify immunological findings to specific behavioral symptoms such as clinical behavioral regression. Large cohort studies on well defined patient and aged matched control groups will be needed to unravel the importance of the immune system in autism.

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